

## Automated Extraction of Nucleic Acid from Serum Samples with the KingFisher™ Flex

### Materials provided

Contents	Cat. No. TC-9119-480 (480 reactions)	Storage
VetAlert™ MagBead Proteinase K	25 mL	15-30°C (room temperature)
VetAlert™ MagBead Lysis Solution	190 mL	
VetAlert™ MagBead Wash Buffer	485 mL	
VetAlert™ MagBead Elution Buffer	45 mL	
VetAlert™ MagBead Resin	22 mL	
VetAlert™ MagBead Stabilization Buffer*	100 mL	
VetAlert™ MagBead Stabilization Additive*	2 mL	2-8°C (refrigerated)

\*Included in kit but not used in this protocol

### Materials required but not provided

#### Reagents

- 100% ethanol, ACS reagent grade or equivalent
- 100% isopropanol, ACS reagent grade or equivalent
- Molecular biology grade water
- (Optional) VetAlert™ DNA Internal Control System [Tetracore, Inc. Cat.# TC-9123-100, includes VetAlert™ DNA Internal Control (DNA IC) and VetAlert™ Internal Control Primer Probe Mix]
- (Optional) VetAlert™ RNA Internal Control System [Tetracore, Inc. Cat.# TC-9122-100, includes VetAlert™ RNA Internal Control (RNA IC) and VetAlert™ Internal Control Primer Probe Mix]

#### Equipment and consumables

- KingFisher™ Flex Purification System (Thermo Fisher Cat.# 5400630)
- Eppendorf ThermoMixer® C (Eppendorf Cat.# 5382000023) or similar and compatible microcentrifuge tubes
- Micropipettes and sterile pipette tips with aerosol barriers
- Vortex mixer
- Microcentrifuge
- KingFisher™ 96 deep-well plates (Thermo Fisher Cat.# 95040460)
- KingFisher™ 96 well microplate, 200 µL (Thermo Fisher Cat.# 97002540)
- KingFisher™ 96 tip comb for deep-well magnets (Thermo Fisher Cat.# 97002534)

## Before you begin

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### Prepare 50% Ethanol:

- Dilute 100% ethanol to 50% with an equal volume of molecular biology grade water.

### Preprocessing of samples

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1. Transfer 250  $\mu$ L of sample to a microcentrifuge tube.
2. Add 350  $\mu$ L of Lysis Solution and 30  $\mu$ L of Proteinase K to each sample.  
  
**Optional:** Add 6  $\mu$ L of VetAlert™ RNA IC or VetAlert™ DNA IC to the Lysis Solution if IC is used as an extraction control.
3. Mix sample on an Eppendorf Thermomixer (or similar) at 56°C and 1400 rpm for 15 minutes.
4. Allow sample to cool at room temperature for 5 minutes.
5. Briefly centrifuge the sample tubes before preparing the Sample Plate (Step 10).

### Prepare purification plates and sample plate

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6. **Wash Plate 1:** Add 100  $\mu$ L of 50% ethanol and 900  $\mu$ L of Wash Buffer to each well of a KingFisher™ 96 deep-well plate required for purification.
7. **Wash Plate 2:** Add 450  $\mu$ L of 50% ethanol to each well of a KingFisher™ 96 deep-well plate required for purification.
8. **Elution Plate:** Add 50  $\mu$ L of Elution Buffer to each well of a KingFisher™ 96 well microplate (200  $\mu$ L) required for purification.
9. **Tip Comb Plate:** Place KingFisher™ 96 tip comb into an empty KingFisher™ 96 well microplate (200  $\mu$ L).
10. **Sample Plate:**
  - a. Transfer 600  $\mu$ L of sample lysate to a well of a KingFisher™ 96 deep-well plate.
  - b. Add 350  $\mu$ L of 100% isopropanol to each well containing sample.
  - c. Add 20  $\mu$ L of Resin to each well containing sample.

**Note:** The magnetic beads tend to drop out of suspension when not in use. Please ensure the Resin is completely resuspended prior to transfer by vigorously shaking or vortexing the bottle.

**Begin process on the KingFisher Flex**

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11. Start KingFisher method *Tetracore\_VetAlert\_MagBeadExtraction.bdz* and follow instrument prompts for plate placement (see below).

Plate ID	Plate position on the KingFisher Flex
Sample Plate	1
Wash Plate 1	2
Wash Plate 2	3
Elution Plate	4
Tip Comb Plate	5

12. Follow prompts for removal of plates from the KingFisher Flex.
13. Transfer final eluates to a container of your choice for storage.

**Note:** Extracted nucleic acid can be stored at 4°C (2°C to 8°C) for immediate use, or at -20°C (-15°C to -25°C) or -80°C (-60°C to -90°C) for long-term storage.

### Symbols

Symbol	Meaning	Symbol	Meaning
	Catalog Number		Lot Number
	Consult Instructions for Use		Manufacturer
	Expiration Date		Temperature Limit

### Revision History

Revision	Date	Description of Change
00	18 July 2024	Initial Release

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